## Chemical composition of the Rotifer (*Brachionus plicatilis*, Muller) fed on *Brachiomonas* sp. and *Eunotia* sp.

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As a live food, rotifer (<u>Brachionus plicatilis</u> Muller) is well-suited to the purpose of rearing the larvae of most marine fish, due to its appropriate size, rapid production rate and capability to be fed on a variety of live unicellular alges or baker's yeast. Nutritional quality of the rotifer is very important in the survival of fish larvae (Howell, 1977). This study aims at investigating the nutritional effects of phytoplankton monocultures <u>Brachiomonas sp.</u> and <u>Eunos</u> which were isolated in Biological Institute in Dubrownik, on the chemical composition of the rotifer. The rotifer's sumples were taken at exponential, stationary and death phases in order to determine water, ash, total lipids, proteins and carbohydrates contents

Algae were cultured in the pasteurized natural sea-water enriched with nutrient and in their late exponential phase of growth, the rotifer was added. The rotifers were separated on 53 µm aperture naylon mesh. In the samples the moisture were determined by drying at 60° C and ash content by ashing at 800° C (Lovergrove, 1966). Lipids were extracted with a chloroform-methanol mixture and estimated by the sulphophospho-vanillin method (Barnes and Blackstock, 1973). To the lipid free pellets TCA was added. In supernatant total carbohydrates determinations were done using a phenol sulfuric acid method reported by Kochert (1978). In precipitate protein was assayed as described by Bradford (1976). Three experimental series were performed.

Table I shows the values of water, ash, lipids, carbohydrates and proteins content in rotifers fed on the green algae <u>Brachiomonas sp.</u> and the diatoms <u>Bunotia</u> <u>Bp.</u> at exponential, stationary and death phases. In the both samples water content reached its lowest value at the stationary phase of growth. Ash levels were highest at the death phase, probably due to a decline in organic matter. Its highest value was found in <u>Bunotia sp.</u> fed rotifer and probably was consistent with siliceous nature of cell walls of diatoms. Lipids and carbohydrates were observed to decline from the exponential to the death phases. Protein levels increased at the stationary phase and reached the highest value in <u>Bunotia spi</u>-fed rotifers. At the last growth phase a marked and faat decline in lipids, carbohydrates and proteins was observed along with an increase in ash and water contents. The above results indicate that the rotifer should be maintained at the late exponential phase when it was observed to be most suitable to the feeding purposes.

In our further research rotifers at the late exponential phase of growth fed on <u>Brachiomonas sp.</u> and <u>Runotis sp.</u> should be used as a diet for fish larvae. The chemical composition and the survival rate of fish larvae would be observed.

TABLE I: Moisture (%wet weight), ash, lipid, carbohydrat, protein (%dry weight), Growthcycle: I-exponential, II-stationary and III-death phases. Means at the same phase of growth followed by different superscripts are significantly different (PK0,05, Student's t-test) Inoculum on Day 0 contained 89,7% moisture, 6,9% ash, 12,4% lipid, 2,1% carbohydrat and 38,4% protein.

ROTIFERS FED ON							
Pt GI	HASES OF	BRACHIOMONA SD.	s eunotia so	BRACHIOMONAS SQ	EUNOTIA SD.	HASES	оғ н
	I	13,9°	14,5°	87,5°	89,7	1	
	Π	11,5°	13,3°	86,9ª	88,3ª	$\Pi$	MOISTURE
	Ш	10,3ª	12,2ª	90,0ª	91,7°	I	
Carbo- Hydrat-	I	3,5°	4,1ª	6,1ª	10,2 <sup>b</sup>	I	
	I	2,9ª	3,8°	7,9ª	12,7°	1	ASH.
	Ш	2,2ª	2,8ª	13,2°	17,0°	$\Pi$	
PROTEIN	I	28,7ª	42,3 <sup>▶</sup>				
	I	45,1°	54,6				
	II	34,2°	38,5°				

## REFERENCES.

BARNES, H. and BLACKSTOCK, J., 1973. Estimation of lipids in marine animals and tissues: detailed investigation of the sulphophospho-vanillin method for total lipids. J. Exp. mar. Biol. Ecol., 12: 103-118.

BRADFORD, M. M., 1976. A rapid and sensitive method for the quantitation of micrgram quantities of protein utilizing the principle of protein-dye binding. Anal. Biochem., 72: 248-254.

BOWELL, B. R., 1977. Aspects of the development of cultivation techniques for flatfish, Ph. D. Thesis, M.A.F.F., Fisheries Laboratory, Lowestoft U.K., 53: 833-838.

KOCHERT, G., 1978. Carbohydrate determination by the phenol-sulfuric acid method. In: Handbook of phycological methods: physiological and biochemical methods. Ed. by J. A. Hellebust and J.S. Craigie. London: Cambrige University Press: 95-97.

LOVERGROVE, T., 1966. The determination of the dry weight of plankton and the effect of various factors on the values obtained. In: Some contemporary studies in marine science. Ed. by H. Barnes, London: 429-467.

Rapp. Comm. int. Mer Médit., 32, 1 (1990).