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The sediment-water interface is characterized by high biological activity and steep concentration gradients of many chemical species. Ordinary chemical analysis in these steep gradients is often impossible to perform with sufficient depth resolution, and the only alternative is to use microsensors. Presently, electrochemical microsensors for O₂, pH, N₂O, dissolved sulfide, and redox potential have been used in marine research, and in fresh water environments it is furthermore possible to analyze NO₃⁻ and NH₄⁺. Work is currently being conducted to expand the arsenal with a microsensor for dissolved inorganic carbon (CO₂ or bicarbonate). In addition to the chemical sensors fiber-optic light sensors for irradiance and scalar irradiance have been developed. Chemical microsensors with optical signal detection (optrodes) have been developed, and such optical sensors may in the future significantly expand our arsenal of chemical species which can be analyzed.

The analysis of by use of microsensors may be performed on sediment cores brought to the laboratory, but it is also possible to do in situ analysis. Simple in situ analysis at shallow water depth can be done with laboratory equipment which is brought to the field, but analysis in the deep sea requires sophisticated computerized equipment. Jens Gundersen and Bo Barker Jorgensen in our group have developed a lander which can be used down to 6000 m water depth and which can record 8 simultaneous microprofiles at 25 μm depth resolution. The microsensors which have been used until now are those for O₂, pH, and dissolved sulfide. The microsensors are identical to those used in the laboratory except that they are pressure compensated by a flexible rubber bulb filled with paraffin oil. Even in the deep sea, profiles can be recorded with extreme accuracy in terms of both concentration and spatial resolution. Among the interesting results obtained from the deep sea work is the finding that the analysis of near-interface pore water chemistry on sediment cores brought to the surface and exposed to atmospheric pressure will result in very erroneous results.

Sediments from shallow depths are exposed to daylight, and the photosynthetic activity by microorganisms within the uppermost sediment layers very much affects the pore water chemistry and thereby all microbial metabolism. We have previously shown how photosynthesis in sediments by use of microsensors can be analyzed at 0.1 mm depth resolution. The transformation and assimilation of inorganic nitrogen species were studied in various types of biofilms and sediments using ¹⁵N and microsensor methods. By use of microsensors for O₂, N₂O, and NO₃⁻ it was possible to localize the depth horizons active in nitrification and denitrification, and it was also possible to obtain depth profiles of reaction rates. Nitrification occurred in the oxic surface layer, and in dark incubated sediments it was sometimes possible to distinguish two distinct nitrification maxima where the upper one located near the sediment surface was based on NH₄⁺ from the waterphase and the lower one just above the oxic-anoxic interface was based on diffusion from deeper sediment layers. Some of the formed NO₃⁻ diffused out of the sediment (uncoupled nitrification) whereas the rest diffused down into anoxic layers where it was denitrified (coupled nitrification-denitrification). Denitrification was always restricted to the deeper anoxic or almost anoxic layers where NO₃⁻ was the most favorable electron acceptor present. The denitrification zone could be extremely thin (< 100 μm) at low NO₃⁻ concentrations in the overlying water and at low nitrification rates, but could also be several millimeters thick at high NO₃⁻. The concentration of NO₃⁻ in any specific layer was not limiting denitrification as long as more than ca. 10 μM NO₃⁻ was present. Photosynthetic biofilms exhibited pronounced diurnal cycles in assimilation of NO₃⁻, nitrification, and denitrification. Denitrification in the photic zone stopped when the sediment was illuminated and oxygen was produced by photosynthesis, but denitrification started immediately again after darkening and onset of anoxic conditions. A new ¹⁵N isotope method, which is based on monitoring the frequency of ¹⁵N₂, ¹⁴¹⁵N₂, and ¹⁴¹⁴N₂ in N₂ gas evolved after adding ¹⁵NO₃⁻ to the overlying water, was used to determine rates of nitrification and denitrification. Determination of total nitrification rates was only possible when also the isotope dilution in waterphase NO₃⁻ was monitored. This new isotope technique has for the first time enabled us to do determinations of nitrification and denitrification (including coupled nitrification-denitrification) in all types of biofilms and sediments without the addition of artifact-creating artificial inhibitors of metabolic processes. Investigations of a thick photosynthetic biofilm showed that coupled nitrification-denitrification was two times higher during illumination than during darkness due to better oxygen conditions for nitrification. Denitrification based on NO₃⁻ in the overlying water was reduced to about half during illumination due to the longer diffusion path to the denitrifying anoxic layers. Assimilation by the microflora was high in the light and remained high for many hours after darkening. This high rate of dark NO₃⁻ assimilation has previously erroneously been interpreted as dissimilatory reduction to NH₄⁺ as mediated by strictly anaerobic bacteria.

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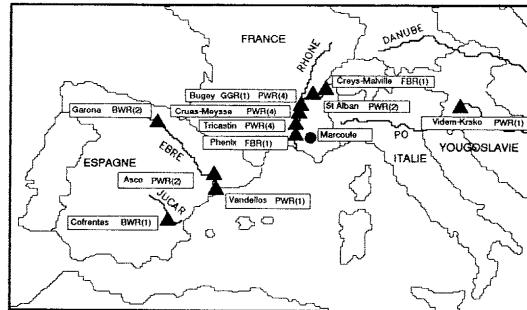
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Lors de la réunion du Comité de Radioactivité Marine durant le Congrès Assemblée plénière de la CIESM à Athènes (1988) il a été décidé de promouvoir un programme sur le thème : Inventaire Global de la Radioactivité de la Mer Méditerranée (Mer Noire et détroit de Gibraltar y compris) ayant pour sigle GIRMED. Un tel programme est apparu essentiel pour répondre aux recommandations de la Convention de Barcelone pour la protection de la mer Méditerranée contre la pollution chronique ainsi que pour l'évaluation des conséquences d'une éventuelle contamination accidentelle de l'environnement marin comme ce fut le cas après l'accident de Tchernobyl.

Les principaux objectifs de GIRMED sont :

- l'intercalibration des mesures réalisées dans les laboratoires impliqués dans des programmes nationaux de surveillance radiologique,
- la cartographie spatio-temporelle des niveaux de radioactivité des principaux radioéléments artificiels dans l'eau, les sédiments et les organismes marins,
- la sélection de bioindicateurs représentatifs des niveaux de radioactivité,
- l'étude des transferts des radioéléments aux populations humaines via des composantes marines, et
- l'évaluation des coefficients de partage entre l'eau, les sédiments et les organismes marins.

Ce projet repose sur la collaboration de 15 laboratoires de 8 pays méditerranéens. De 1988 à 1992 ces laboratoires ont participé à un programme d'assurance qualité de leur mesures par l'intermédiaire d'exercices d'intercalibration. En parallèle l'inventaire des sources de rejets de radioéléments dans l'environnement marin méditerranéen a été réalisé et une base de données des mesures réalisées dans l'environnement a été mise en place. La deuxième phase conduira à l'estimation des doses pour les populations concernées par les sources de radioactivité en Méditerranée.



Centrales nucléaires (▲) et centre de retraitement (●) au 1/1/91 (bassin occidental).

EAU DE MER

Pays	Lieu	Date	Alp	Ru	Cs	Ag	Sb	Ce
FRANCE	côtes	4/86 à 9/87		X	X		X	
MONACO	côtes	5/86 à 6/86			X	X		
ROUMANIE	Estuaire Danube	10/87				X		

SEDIMENT MARIN

Pays	Lieu	Date	Alp	Ru	Cs	Ag	Sb	Ce
FRANCE	côtes	11/86 à 3/91	X	X	X	X	X	X
MONACO	Cannes	2/87						
GREECE	Milos	12/86		X		X		
ROUMANIE	côtes	5/86 à 11/90	X			X		
TURQUIE	Akkuyu	6/89				X		

ALGUES

Pays	Lieu	Date	Alp	Ru	Cs	Ag	Sb	Ce
TURQUIE	Noire	7/86 à 1/89		X	X			
ROUMANIE	côtes	10/86 à 7/90	X	X	X	X		X
FRANCE	côtes	6/86 à 9/87		X	X	X	X	X
MONACO	côtes	1/86 à 2/87		X	X	X		
GREECE	Milos	12/86	X			X		
ROUMANIE	côtes	7/90	X			X		
TURQUIE	Akkuyu	6/89				X		
ROUMANIE	Noire	7/86 à 6/89		X	X			

POSITIONS

Pays	Lieu	Date	Alp	Ru	Cs	Ag	Sb	Ce
FRANCE	côtes	1/86 à 6/91		X	X	X	X	X
GREECE	Milos	12/86	X		X			

MATIERE EN SUSPENSION

Pays	Lieu	Date	Alp	Ru	Cs	Ag	Sb	Ce
FRANCE	rhône	9 à 3/87		X	X	X		
MONACO	Monaco offsh	4/86 à 6/86	X	X	X			
ITALIE	La Spezia	7/86		X	X			

MOLLUSQUES BIVALVES

Pays	Lieu	Date	Alp	Ru	Cs	Ag	Sb	Ce
ROUMANIE	côtes	6/87 à 6/89	X	X	X			
FRANCE	côtes	1/83 à 9/91	X	X	X	X	X	X
MONACO	côtes	5/86 à 12/86		X	X	X	X	X
ROUMANIE	côtes	8/86 à 5/90	X	X	X			
TURQUIE	Bosphore	5/86 à 2/87		X	X			
	Noire	2/87 à 2/88		X	X	X		

POISSONS

Pays	Lieu	Date	Alp	Ru	Cs	Ag	Sb	Ce
FRANCE	Thau	3/86 à 9/88		X	X	X	X	X
	Beauduc	2/86 à 8/88		X	X	X	X	X
	Nice	6/86 à 9/91		X	X	X	X	X
TURQUIE	Marmara	5/86 à 1/87		X	X			
	Noire	12/86 à 1/87		X	X			
	Bosphore	5/86		X	X			
GREECE	Milos	12/86	X		X			
ROUMANIE	Constanta	6/87 à 6/89		X				

Etat de la base de données GIRMED. Classement par type d'échantillons et pays pour la période 1986-1991.
Alp : émetteurs alpha soit ²³⁹Pu, ²³⁸Pu, ²³⁸Ra, ²²⁶Ra, ²²⁸Ra, ²³²Ra; Ru : ¹⁰³Ru; Cs : ¹³⁴Cs et ¹³⁷Cs; Ag : ¹¹⁰Ag; Sb : ¹²⁵Sb; Ce : ¹⁴⁷Ce et ¹⁴⁴Ce; les données sur les Polychètes Melina ne figurent pas dans ce tableau.