PHOSPHOLIPASE A2 IN MARINE INVERTEBRATES

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Abstract

Phospholipase A2 (PLA2) catalytic activity was determined by a radiochemical assay in tissue extracts from 21 species belonging to five phyla of marine invertebrates that were collected from the Great Barrier Reef, Queensland, Australia. The highest PLA2 activities were found in hard coral, fire coral, crown-of thorns starfish and sea cucumber. High PLA2 activities were found in a number of echinoderms and sea anemones, whereas annelids, crustaceans and molluscs contained relatively low amounts of PLA2 activity. The results demonstrate the presence of PLA2 activity in a number of marine invertebrates. The molecular structure, classification and physiological functions of these PLA2s remain to be studied.

on whole animal extract.

Soft coral Sinularia flexibilis

Soft coral Sarcophyton elegans Soft coral Dendronephthya sp.

Cnidaria

Key words : Cnidaria, echinodermata, mollusca, toxins

Introduction

Phospholipases A2 (PLA2) form a large family of lipolytic enzymes (1). PLA2 is a major component of snake and other venoms, digestive secretions of the gastrointestinal tract, as well as secretions of various mucous surfaces including tears and seminal fluid. In addition to toxic and digestive functions, PLA2 has effective bactericidal properties and participates in the regulation of inflammation by releasing arachidonic acids from cellular membrane phospholipids for eicosanoid synthesis. The purpose of the current study was to investigate the occurrence of PLA2 in marine invertebrate tissues

Material and methods

Specimens were collected at 0-20 m depth from the Great Barrier Reef, Northern Oueensland, Australia, Samples were immersed in 50 mM acetate buffer, pH 5, containing protease inhibitors, and frozen at -18°C. After thawing, the specimens were homogenised by Ultra Turrax or shaken vigorously in a glass container (coral specimens) and centrifuged at 5 000 g. The supernatants were assayed for PLA2 activity by using 14Clabelled phosphatidylcholine in mixed micelles as a substrate. Protein was determined by a standard dye-binding assay.

Results and discussion

The highest PLA2 activities were found in the extracts of hard coral, fire coral, crown-of thorns starfish and sea cucumber (Table 1). The high PLA2 content in the puffer fish intestine is similar to mammalian intestine where the enzyme is expressed in mucosal Paneth cells (2). High PLA2 activities were found in a number of echinoderms and sea anemones, whereas the annelids, crustaceans and molluscs tested had relatively low PLA2 activity. The functions of PLA2 in invertebrates are not well known. The action of PLA2 on phospholipids initiates the synthesis of eicosanoids present in most animal species including invertebrates (3). Insect immune response to bacteria is mediated by eicosanoids (4). PLA2 activity has been reported in the granular amebocytes, important immunocompetent cells of the horseshoe crab, Limulus polyphemus (5). PLA2 is a well-characterised digestive enzyme in mammals (1). Digestive PLA2s have been found in the tiger beetle Cicindella circumpicta (6) and the starfish Asterina pectinifera (7). The presence of PLA2 in snake venoms has been known since the 1890' (1) and has been reported in numerous invertebrate venoms including that of the marine snail Conus magus (8), the scorpion Pandinus imperator (9), the ant Pseudomyrmex triplarinus (10), the sea anemone Aiptasia pallida (11) and the jellyfish Rhopilema nomadica (12). It is pertinent that the highest levels of PLA2 observed in echinoderms in the present study were found in tissues that are associated with toxins; E.G. the spines of the crown-of-thorns starfish have a powerful neurotoxin. Pyloric cecae of echinoderms contain PLA2 that may function as a digestive enzyme.

The current results demonstrate the presence of PLA2 activity in a number of marine invertebrates. The molecular structure, classification and physiological functions of these PLA2s remain to be studied.

References

1- Six D.A. and Dennis E.A., 2000. Review. The expanding superfamily of phospholipase A2 enzymes: classification and characterization. Biochim Biophys. Acta, 1488: 1-19.

2- Nevalainen T.J., Grönroos J.M. and Kallajoki M., 1995. Expression of group II phospholipase A2 in human gastrointestinal tract. Lab. Invest. 72: 201-208

3- Stanley-Samuelson D.W., 1991. Comparative eicosanoid physiology in invertebrate animals. Am. J. Physiol. 260: R849-R853

4- Stanley-Samuelson D.W., Jensen E., Nickerson K.W., Tiebel K., Ogg C.L. and Howard R.W., 1991. Insect immune response to bacterial infection is

mediated by eicosanoids. *Proc. Natl. Acad.* Sci. USA 88: 1064-1068. 5- McPherson J.C. and Jacobs R.S. An 18.5 kDa protein from the amebocyte of Limulus polyphemus, homologous to the previously described amebocyte aggregating factor, expresses alternative phospholipase A2 activity. Comp. Biochem. Physiol. B, 127: 31-44.

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Hard coral Acropora sp. 267

Table 1. Phospholipase A2 activity concentration (U/g protein) in tissue extracts. Where a specific tissue is not indicated the assay was undertaken

U/a (SD)

22 (6) 82 (91) 64 (12)

n

2 4 10

1

Hard coral Pocillopora damicornis	945 (540)	10
Hydrozoan fire coral Millepora sp.	/ 35	1
Sea anemone Stoicnactis sp.	207 (35)	2
Sea anemone Actinia australis	90 (24)	2
Annelida	20 (22)	4
worm Phyliodoce novaenollandiae	29 (32)	4
Crustacea	2 (2)	7
Gnost crab Ocypode cordinana	3 (2)	/
Prawn Panaeus monodon	EQ (70)	2
Hepatopancreas	52(79)	3
Mallusse	0.5 (0.2)	2
Nollusca	4	4
Clem Deney supportue	1 2 (1)	i c
Viam Donax curreatus	2(1)	1
Rudibanch Phyllida Sp.	00	1
Crinoid Coloboratro poroninoso	124	1
Brittle stor Ophiosome cripsocus	124	
Dinie star Ophiocoma enhaceus	22	1
DISC Arm	20	1
Starfish Comonbia sp	25	1
Starfish Fromis on	200	1
Starfish Lingkia loguigata	200	1
Crown of thorns starfish Aconthestor planci	244	1
Skin	97	1
Snings	1625	1
Body wall	117	1
Pyloric ceca	498	1
Sea cucumber Stichonus chloronotus	100	
Body wall	5423	1
Intestine	235	1
Rete mirabile	614	1
Vertebrata	• • •	
Pufferfish Arothron manilensis		
Skin	177 (87)	5
Muscle	4 (2)	5
Liver	78 (73)	5
Intestine	2753 (1058)	5
	/	2

6- Uscian J.M., Miller J.S., Sarath G. and Stanley-Samuelson D.W., 1995. A digestive phospholipase A2 in the tiger beetle *Cicindella circumpicta*. J. Insect Physiol., 41: 135-141.

7- Kishimura H., Ojima T., Hayashi K. and Nishita K., 2000. cDNA cloning and sequencing of phospholipase A2 from the pyloric ceca of the starfish Asterina pectinifera. Comp. Biochem. Physiol. B, 126: 579-586.

8- McIntosh J.M., Ghomashchi F., Gelb M.H., Dooley D.J., Stochr S.J., Giordani A.B., Naisbitt S.R. and Olivera B.M., 1995. Conodipine-M, a novel

phospholipase A2 isolated from the venom of the marine snail Conus magus. J. Biol. Chem. 270: 3518-3526.

9- Zamudio F.Z., Conde R., Arévalo C., Becerril B., Martin B.M., Valdivia H.H. and Possani L.D., 1997. The mechanism of inhibition of ryanodine receptor channels by imperatoxin I, a heterodimer protein from the scorpion Pandinus imperator. J. Biol. Chem. 272: 11886-11894.

10- Hinks W.F., Pappas P.W. and Jaworski D.C., 1994. Partial biochemical characterization of venom from the ant, Pseudomyrmex triplarinus. Toxicon 32:763-772

11- Grotendorst G.R. and Hessiger D.A., 2000. Enzymatic characterization of the major phospholipase A2 component of sea anemone (*Aiptasia pallida*) nematocyst venom. *Toxicon* 38: 931-943. 12- Gusmani L., Avian M., Galil B., Patriarca P. and Rottini G., 1997.

Bologically active polypeptides in the venom of the jellyfish Rhopilema nomadica. Toxicon 35: 637-648.