

# INFLUENCE OF SMALL INCREASES IN TEMPERATURE ON PLANKTONIC BACTERIAL CARBON USE IN THE BLANES BAY (COASTAL NW MEDITERRANEAN)

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## Abstract

Bacterial production (BP) and respiration (BR) were measured seasonally in a Mediterranean coastal area (Blanes Bay) at two temperatures: ambient temperature (TA) and about two degrees above that temperature (T+2). Seawater samples were filtered through 0.8  $\mu\text{m}$  to obtain a predator-free bacterioplankton assemblage and incubated for 48 hours at both temperatures. From production and respiration rates we derived bacterial growth efficiency (BGE). Production and respiration rates were positively affected by the increase in temperature, while BGE remained unchanged. Therefore, small increases of temperature might increase the amount of carbon passing through bacteria.

**Keywords:** Bacterial production. Bacterial respiration. Bacterial growth efficiency. Temperature.

## Introduction

Due to human industrialization and deforestation the concentration of  $\text{CO}_2$  has increased in the atmosphere [1], and this is expected to lead to an increase in Earth's average temperature of  $\sim 2.5$   $^\circ\text{C}$  in a century [2]. Temperature has an extremely important influence on biological processes such as bacterial production and respiration [3, 4]. As microbes are relevant actors in the transfer of carbon in aquatic ecosystems [5], there is an increasing interest in the effect of small temperature increases on microbial plankton [6]. The objective of this study is to test the effect of small increases in temperature (ca.  $2^\circ\text{C}$ ) on bacterial carbon use and growth efficiency in an oligotrophic coastal system (NW Mediterranean) and its implications for the planktonic carbon cycle.

## Methodology

Experiments were done monthly (March-August 2003) with subsurface seawater collected from a fixed station in Blanes bay ( $41^\circ 39' \text{N}$ ,  $2^\circ 48' \text{E}$ ), over 15 m depth and about half mile offshore. Seawater temperature was determined with a calibrated thermometer, and 25 liters were pre-filtered through 200  $\mu\text{m}$ , placed on a carboy, and transported to the laboratory. The samples were then filtered through 0.8  $\mu\text{m}$  (AAWP, Millipore), bacterial production rates determined by  $^3\text{H}$ -Leucine uptake [8] and converted to bacterial carbon with a standard factor of 3.1  $\text{Kg C mol}^{-1}$ . Filtered water was also distributed in 48 borosilicate glass bottles (ca. 130 ml), of which 8 were immediately fixed with Winkler reagents (t0). The remaining bottles were placed in two temperature-controlled chambers, set at ambient temperature (TA) and  $\sim 2$  degrees above this temperature (T+2). At 24 and 48 hours we fixed the remaining 40 bottles, 10 each time for each temperature. Dissolved oxygen was determined with an automatic titrator based on potentiometric endpoint detection [7]. Respiration rates were obtained by linear regression of oxygen concentration vs. time, and were transformed to carbon units assuming a respiration quotient of 1. Bacterial carbon use is  $\text{BP} + \text{BR}$  and bacterial growth efficiency (BGE) is  $\text{BP}/(\text{BP} + \text{BR})$  [9].

## Results and discussion

Seawater temperature varied twelve degrees in Blanes bay, between  $13$   $^\circ\text{C}$  in March and  $25$   $^\circ\text{C}$  in August. Initial bacterial productions varied between  $1.3$   $\mu\text{g C l}^{-1} \text{d}^{-1}$  in June and  $150$   $\mu\text{g C l}^{-1} \text{d}^{-1}$  in July, increasing exponentially at 24 hours during the incubations and showing a clear shift-up in the T+2 samples (Fig. 1). The average increase of bacterial production ( $(\text{BP}_{\text{TA}} - \text{BP}_{\text{T+2}})/\text{BP}_{\text{TA}}$ ) was  $36$  ( $\pm 12$ ) %. BP's at both temperatures were positively correlated,  $\text{BP}_{\text{T+2}} = 15.3$  ( $\pm 20.3$ ) +  $1.1$  ( $\pm 0.1$ )  $\text{BP}_{\text{TA}}$  ( $n = 18$ ,  $r^2 = 0.8$ ,  $p < 0.01$ ). Respiration rates at ambient temperatures varied between  $9.2$   $\mu\text{g C l}^{-1} \text{d}^{-1}$  in May and  $102.1$   $\mu\text{g C l}^{-1} \text{d}^{-1}$  in March. The average increase of respiration rates at the higher temperature was  $27$  ( $\pm 11$ ) %. Both respiration rates were significantly related,  $\text{BR}_{\text{T+2}} = 3.2$  ( $\pm 2.7$ ) +  $1.1$  ( $\pm 0.05$ )  $\text{BR}_{\text{TA}}$  ( $n = 18$ ,  $r^2 = 0.99$ ,  $p < 0.01$ ). Finally, bacterial growth efficiencies varied between 4.7 and 95 %, and were not influenced by the small increase in temperature,  $\text{BGE}_{\text{T+2}} = 4.5$  ( $\pm 3.7$ ) +  $0.95$  ( $\pm 0.05$ )  $\text{BGE}_{\text{TA}}$  ( $n = 18$ ,  $r^2 = 0.95$ ,  $p < 0.01$ ). Adding the values up, it can be calculated that a small temperature increase could enhance bacterial carbon use by near a 60 %.

We obtained experimental data that agree with modeling studies in which small increases in temperature had a positive effect on bacterial production, respiration and carbon demand [3, 4], but no influence on BGE [9]. Therefore, small changes in temperature will have an effect on the microbial components of aquatic systems [6] significantly increasing the amount of carbon processed by bacteria.

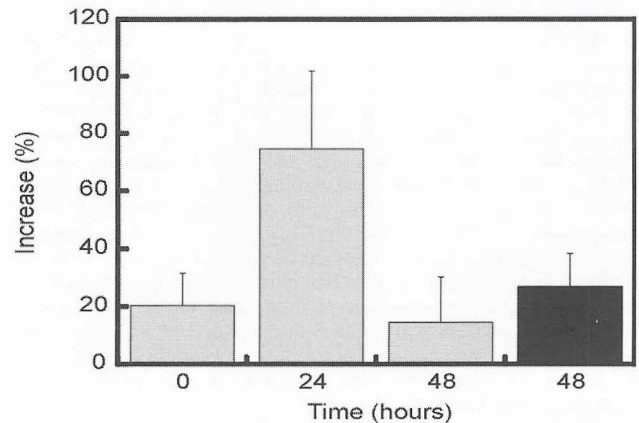


Fig. 1. Average increase in BP and BR at time 0, 24 and 48 hours (BP, gray columns) and during 48 hours (BR, black column).

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## References

- 1 - Siegentaler U. and Sarmiento J.L. Atmospheric carbon dioxide and the ocean. *Nature*, 365: 119-125.
- 2 - Houghton J., 1997. Global warming: the complete briefing. 251 pp. In: Houghton J. (ed.), Cambridge University Press. Cambridge, U.K.
- 3 - Pomeroy L.R. and Wiebe W.J., 2001. Temperature and substrates as interactive limiting factors for marine heterotrophic bacteria. *Aquat. Microb. Ecol.*, 23: 187-204.
- 4 - Rivkin R.B. and Legendre L., 2001. Biogenic carbon cycling in the upper ocean: Effects of microbial respiration. *Science*, 291: 2398-2400.
- 5 - Azam F., Fenchel T., Field J.G., Gray J.S., Meyer-Reil L.A., Thingstad F., 1983. The Ecological Role of Water-column Microbes in the Sea. *Mar. Ecol. Prog. Ser.*, 10: 257-263
- 6 - Petchey O.L., McPhearson P.T., Casey T.M., Morin P.J., 1999. Environmental warming alters food-web structure and ecosystem function. *Nature*, 402: 69-72.
- 7 - JGOFS. (Joint Global Flux Study) 1994. Protocols for the JGOFS core measurement. In: Knap A., Michaels A., Cloze A., Ducklow H., Dickson A. (eds.) JGOFS Rep. No 19.
- 8 - Smith D.C. and Azam F., 1992. A simple, economical method for measuring bacterial protein synthesis rates in seawater using  $^3\text{H}$ -leucine. *Marine Microbial Food Webs*, 6: 107-114.
- 9 - del Giorgio P.A. and Cole J.J., Bacterial growth efficiency in natural aquatic systems. *Annu. Rev. Ecol. Syst.*, 29: 503-541.